

Precision in the Detection of Myeloid-derived Suppressor Cells (MDSCs)

Lynn Kisselbach, MSc., Kanjinga Tambwe MSc., Deborah Phippard PhD., Kelly Huang, PhD.

Precision For Medicine, Inc., Frederick, MD

ABSTRACT

Myeloid-derived suppressor cells (MDSCs) are considered a major regulator of the immune response and are a known biomarker associated with responsiveness to CTI A-4- and PD-1-targeting therapies, as well as, a potential target for therapeutic intervention. While there are publications dating back to the early 1970s that describe immune suppressive myeloid cells, MDSCs have only recently been widely appreciated due to a greater understanding of the phenotype of this heterogeneous population of cells. Recent efforts to harmonize flow cytometric detection of MDSCs across labs internationally led to the following suggestion on the human MDSC phenotype for monocyte-MDCSs (M-MDSCs) as CD11b+CD14+HLA DR-/loCD15⁻, polymorphonuclear-MDSCs (PMN-MDSCs) as CD11b+CD14-CD15+, and early-MDSCs (eMDSCs) as Lin-CD14-HLA-DR-/loCD33+. The objective of this study was to develop and validate a CLIA 7-color flow cytometry assay performed on a BD FACSCanto IVD to reliably detect these MDSC populations in peripheral blood mononuclear cell (PBMC) preparations for use in clinical studies Evaluation of PBMCs isolated using density gradient separation and SepMate[™] tubes was chosen, as this is currently the best method to enrich for PMN-MDSCs, which are part of the low density granulocyte (LDG) population, over normal granulocytes that share surface markers with MDSCs and will confound the analysis.

The MDSC flow cytometry assay met validation criteria for intra-assay, inter-assay, and inter-operator precision, in addition to, specificity, sensitivity/ linearity, and stability. Importantly, this assay clearly demonstrated that the predominant population of MDSCs in stage IV colorectal cancer patients is the PMN-MDSC population and that this PMN-MDSC population is heterogeneous in CD14 expression, with low to negative levels of CD14 expression. Low density granulocytes reportedly can express low levels of CD14; however, it is unknown if this differentiates the PMN-MDSCs from mature pro-tumor neutrophils that are both part of the low density granulocyte population. Monitoring MDSC populations in the clinic with this CLIA assay will support the elucidation of the role MDSCs play in a variety of disease states. We present this biomarker assay as a robust method to stratify patients for multiple immuno-therapies.

PHENOTYPES OF MDSCs: 3 TYPES



Figure 1.

Figure 2.

Phenotype of the three major populations of MDSCs: Monocyte-MDSC (M-MDSC), Polymorphonuclear-MDSCs (PMN-MDSC), and Early MDSC (eMDSC) as reviewed by Bronte et al 2016 Nat Comm.





Step 3. Detection with BD FACSCanto IVD



The first step in this assay is gradient separation of PBMCs from whole blood using the SepMate[™] procedure. This is a key step, as gradient separation is currently the only method to distinguish PMN-MDSCs, found in the low density granulocyte population, from normal granulocytes. Freshly isolated PBMCs either non-stained, stained as an FMO, or full panel stained, as well as, singly stained One Comp Beads[™], were acquired on a BD FACSCanto IVD instrument and the data was analyzed using FlowJo software.



Figure 3.

Assay validation was carried out on cryopreserved healthy donor PBMCs. The table shows the validation parameters tested and the criteria required for passing validation. The top graph shows that upon dilution of fully stained PBMCs into non-stained PBMCs, the assay performs with excellent linearity at the highest dilution of stained cells (25% stained). For the %Total MDSC, % M-MDSC & % PMN-MDSC R² values are 2 0.991, % eMDSC R² value was 0.1297. The bottom graph shows the stained cells are stable prior to acquisition for at least 44 hours post-staining.

MDSCS IN HEALTHY VS. STAGE IV CRC DONORS

Healthy Donors

| Population | M-MDSC | PMN-MDSC | eMDSC | Total circulating MDSC in PBMC |
|------------|--------|----------|----------|-------------------------------------|
| Donor ID | % PBMC | % PBMC | % PBMC | % Total circulating MDSC in PBMC |
| Healthy 1 | 2.09 | 0.056 | 1.67E-03 | 2.1 |
| Healthy 2 | 1.83 | 0.11 | 0.49 | 1.7 |
| Healthy 3 | 1.13 | 0.028 | 0.77 | 1.2 |
| Hcalthy 4 | 4.70 | 0.19 | 1.46 | 5.0 |
| Healthy 5 | 1.90 | 0.024 | 1.04 | 2.0 |

Stage IV- Colorectal Cancer (CRC) Donors

| MDSC in PBM | 10 |
|---|----------|
| Donor ID % PBMC % PBMC % PBMC % Total circulat MDSC in PBM | ing C |
| DD17 0.54 0.31 0.92 0.9 | |
| DD18 0.10 5.91 0.65 6.0 | |
| DD19 1.91 71.8 2.22 73.7 | |
| DD20 3.37 44.0 28.4 47.4 | |

Figure 4.

The top table shows the results of the assay performed on 5 healthy donors and the bottom table shows the results of the assay performed on 4 stage IV colorectal cancer patients. All healthy donors had < 10% Total circulating MDSCs (M-MDSCs + PMN-MDSCs); whereas 2 of the 4 stage IV colorectal cancer patients had dramatically higher frequencies of Total circulating MDSCs, primarily driven by the PMN-MDSC population.



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Figure 5. The gating strategy used for data analysis is shown for 1 healthy donor and 2 stage IV colorectal cancer donors, one with and one without elevated Total circulating MDSCs. Clear distinctions were observed in the FSC-A vs SSC-A, CD14 vs CD11b, and CD15 vs SSC-A plots for these donors. Of note, the stage IV CRC cancer donors with the elevated Total circulating MDSC population had an observable CD11b⁺CD14^{Lo} population.

CONCLUSION

This MDSC flow cytometry assay passed all Precision, Specificity, Sensitivity, and Stability validation criteria and therefore can be performed under CLIA regulation. This flow cytometry assay identifies all three MDSC populations and clearly distinguishes between healthy and stage IV colorectal cancer donors. The dominant MDSC population observed in stage IV colorectal cancer patients was the PMN-MDSC population, which was due to an increase in the CD11b+CD14^{Lo/} population.

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Bronte V. et.al., Recommendations for myeloid-derived suppressor cell nomenclature and characterization standards, Nature Communications 7, 12150 (2016).