CHARACTERIZATION INFORMATION

Donor Classification

■ Donor Demographics
  • Gender, Age, Ethnicity

Donor Characterization

■ HLA Typing
  • Class I and Class II
  • High Resolution (A, B and C)
  • High Resolution (DR and DQ)

■ IFNy ELISpot assay results for:
  • PHA, CEF, and CMV Pooled Peptides

INTENDED USE
Cryopreserved Characterized Human Peripheral Blood Mononuclear Cells (PBMCs), Purified are available as controls and ex vivo model systems for cell based assays, flow cytometry assays and immune response monitoring.

For Research Use Only.
Not for use in diagnostic or therapeutic procedures.

PRODUCT DESCRIPTION
Cells are isolated from healthy human donors using a density gradient centrifugation method for purification and minimizing red blood cell content. Cells display ≥80% viability post-thaw. Cells are provided in 1.0 mL or 5.0 mL of freezing media containing 10% DMSO.

Catalog No. 83000C-1.0, Minimum of 10 x 10^6 Cells per Vial
Catalog No. 83000C-5.0, Minimum of 50 x 10^6 Cells per Vial

STORAGE AND HANDLING
All vials of frozen cells should be stored in the vapor phase liquid nitrogen (≤-150°C). When needed for use, transfer up to 4 vials from the vapor phase liquid nitrogen to the lab per the Instructions for Thawing and Culture provided on this page. Vials should be maintained at -70°C or on dry ice for temporary storage and handling up to 4 hours.

PRECAUTIONS
Use Universal Precautions for handling cellular products as for other human specimens.1 Do not pipette by mouth. Avoid direct inhalation of the suspension and handle in areas with adequate ventilation. Do not smoke, eat or drink in areas where specimens are being handled. Dispose of this product as appropriate for biohazardous material.

ORDERING INFORMATION AND TECHNICAL SUPPORT
1-855-222-5010 or buypbmcs@precisionformedicine.com

MANUFACTURED BY
Precision for Medicine
8425 Progress Drive
Frederick, MD 21701

INSTURCTIONS FOR THAWING AND CULTURE
1. Transfer frozen vials to the lab on dry ice to keep them cold. Only thaw 4 vials at one time.
2. Warm media for cell dilution to 37 ± 3°C. For each vial to be thawed, place 10 mL of media in a 50 or 15 mL centrifuge tube.
3. Thaw the frozen vials in water in a 37 ± 3°C water bath with gentle shaking. Just before the last ice crystal has melted, remove the vial from the water. Wipe the vial with a sterile alcohol pad, focusing on the cap area.
4. Add 1 mL of warm culture media from the 50 or 15 mL tube prepared above to the vial using a 1000 µL pipette tip and drop-wise action. Add the culture media over 30 sec to allow the cells to adjust to the change of temperature.
5. Slowly transfer the diluted cells back to the 50 or 15 mL tube. Do not mix or pipette the cells vigorously since cells that have been frozen are initially more sensitive to mechanical stress than fresh cells.
6. Rinse the original vial with 1 mL of the cell containing media from the 50 or 15 mL tube to recover cells that may have adhered to the sides; add the rinse media back to the 50 or 15 mL tube.
7. Pellet the cells by centrifugation at 350xg for 10 minutes with rapid acceleration and brake on. If no pellet is observed, centrifuge at 450xg for an additional 15 minutes. Discard supernatant.
8. Optional Step, Proceed directly to Step 9 if planning to omit DNAse treatment: To avoid any issues with potential cell clumps, resuspend the cell pellet in a small volume of warm media (1-2mL) and mix by gently tapping. Add 100µL of 1mg/mL DNAse I. Mix gently but thoroughly and incubate at room temperature for 15 minutes. Then bring the total volume to the desired counting volume using warm media, mixing the cell solution carefully. Note that if using these optional instructions, DNAse does not need to be washed out of the cells, so proceed directly to Step #11 and perform your cell viability count.
9. Re-suspend the cell pellet by gently tapping and add 10 mL of warm culture media for a second wash. Mix cell solution carefully. Pellet the cells by centrifugation as in step 7.
10. Discard supernatant. Re-suspend the cell pellet by gently tapping and add 5-10 mL of warm culture media.
11. Count cells using laboratory specific procedures and proceed with laboratory protocol assays.

RESTING PERIOD FOR CELLS
Thawed cells may benefit from resting overnight at 37 ± 3°C before use in assays. Adjust viable cell concentration to 2 x 10^6 cell/mL. Put 1-10 mL of cell suspension in a 50 mL conical tube, loosen the tube cap to allow for gaseous exchange, and place the tubes upright in an incubator with the appropriate CO2 level for the media used. After overnight incubation, count and use cells as needed.

PRODUCT SPECIFIC DATA
The characterization information and results for the cells are provided on the lot specific Certificate of Analysis.

REFERENCES
1. CDC recommendations for prevention of HIV transmission in health care settings. MMWR 36 (supp.2) 1987.