

Package Insert

GENERAL INFORMATION

The information below applies to cellular products provided by Precision for Medicine.

INTENDED USE

Cryopreserved Cellular Products are available for use as controls and *ex vivo* model systems for cell based assays, flow cytometry assays and immune response monitoring.

For Research Use Only.

Not for use in diagnostic or therapeutic procedures.

PRODUCT DESCRIPTION

The product descriptions, donor demographics and characterization information are provided on the lot-specific Certificate of Analyses or final reports.

STORAGE AND HANDLING

All vials of frozen cells should be stored in the vapor phase liquid nitrogen ($\leq -150^{\circ}\text{C}$). When needed for use, transfer up to 4 vials from the vapor phase liquid nitrogen to the lab per the Instructions for Thawing and Culture provided on this page. Vials should be maintained at -70°C or on dry ice for temporary storage and handling up to 4 hours.

PRECAUTIONS

Use Universal Precautions for handling cellular products as for other human specimens.¹ Do not pipette by mouth. Avoid direct inhalation of the suspension and handle in areas with adequate ventilation. Do not smoke, eat or drink in areas where specimens are being handled. Dispose of this product as appropriate for biohazardous material.

REFERENCE

1. CDC recommendations for prevention of HIV transmission in health care settings. MMWR 36 (supp.2) 1987.

ORDERING INFORMATION AND TECHNICAL SUPPORT

1-855-222-5010 or buypbmcs@precisionformedicine.com

MANUFACTURED BY

Precision for Medicine
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Frederick, MD 21701

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INSTRUCTIONS FOR THAWING AND CULTURE

1. Warm medium for cell dilution to $37 \pm 3^{\circ}\text{C}$.
2. Transfer frozen vials to the lab on dry ice to keep them cold. Only thaw 4 vials at a time.
3. Hold vials in a $37 \pm 3^{\circ}\text{C}$ water bath. Do not immerse below the level of the cap. Just before the last ice crystal has melted, remove the vial from the water. Wipe the vial with a sterile alcohol pad, focusing on the cap area.
4. Gently pour the contents of the cryovial into a labeled 50 (or 15) mL tube.
5. Add 8 mL of warmed medium dropwise to the 50 mL tube slowly with gentle swirling. This will allow the cells to adjust to the change in environment.
6. Rinse the original vial with 1 mL of warm culture medium and add to the 50 mL tube.
7. Pellet the cells by centrifugation at $400 \times g$ for 10 minutes with rapid acceleration and brake on. If no pellet is observed, centrifuge at $400 \times g$ for an additional 15 minutes. Discard supernatant.
8. **OPTIONAL STEP, PROCEED DIRECTLY TO STEP 9 IF PLANNING TO OMIT DNase TREATMENT:** To avoid any issues with potential cell clumps, resuspend the cell pellet in 2 mL of warm medium containing 0.1 mg/mL DNase I. Mix gently but thoroughly and incubate at room temperature for 15 minutes.
9. Bring cells up to 10 mL using warmed medium then pellet the cells by centrifugation as in step 7.
10. Resuspend cells in the desired volume for counting using warm medium, mixing the cell solution carefully.
11. Count cells using laboratory specific procedures and proceed with laboratory protocol assays.

RESTING PERIOD FOR CELLS

Thawed cells may benefit from resting overnight at $37 \pm 3^{\circ}\text{C}$ before use in assays. Adjust viable cell concentration to 2×10^6 cell/mL. Put 1-10 mL of cell suspension in a 50 mL conical tube, loosen the tube cap to allow for gaseous exchange, and place the tubes upright in an incubator with the appropriate CO_2 level for the media used. After overnight incubation, count and use cells as needed.